

Boosting diagnostics for plant production industries

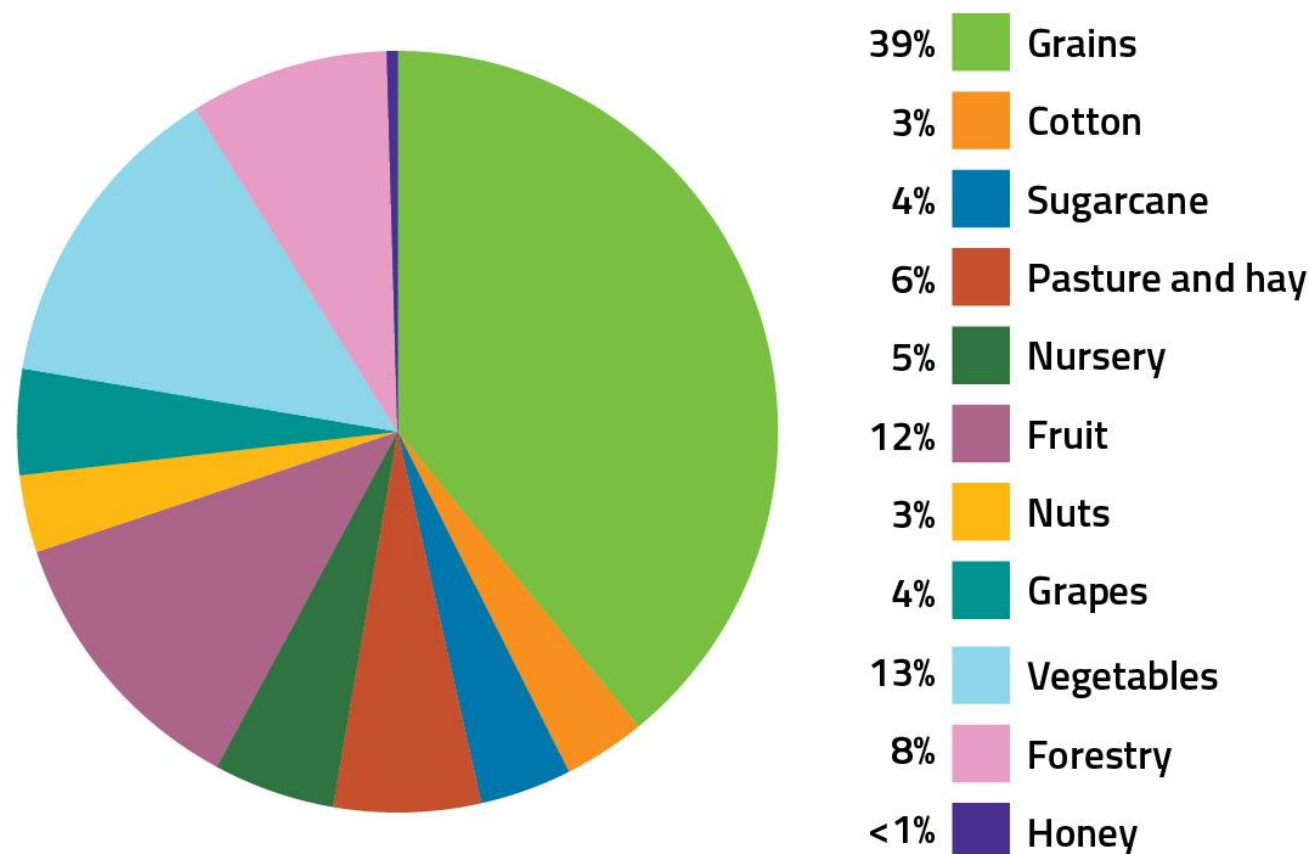
Maddy Quirk
Biosecurity Extension Community

2 March 2022



Solving a general problem

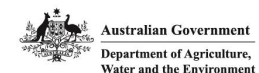
- Plant industries worth approx. \$30b
- Reduced capacity in diagnostics in Australia
- Review conducted on Australia's diagnostic capacity
- Identified areas of improvement needed
- If not addressed, could become a significant issue in future



Contribution of plant production industries to total plant gross value of production in 2017-18 (Source: Plant Health Australia)

Solution: Commonwealth-funded project

- Investment by Department of Agriculture, Water and the Environment's (DAWE) Rural Research and Development for Profit Program funded project – Boosting Diagnostic Capacity for Plant Production Industries
- \$4.6 million from DAWE, \$3 million from RDCs, and \$7 million in-kind.
- 2019 – 2023
- **Objective:** Create maximum value to industry through better partnerships in diagnostic decision-making for biosecurity response efforts.
- Focus areas:
 - Diagnostic capacity building
 - Enhanced diagnostic tools
 - In-field detection and diagnostic blitzes



Complex grouping of R&D projects



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Primary Industries and
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Jobs,
Precincts
and Regions



- Diagnostic capacity building
- Enhanced diagnostic tools
- In-field detection and diagnostic blitzes



Australian Government
Department of Agriculture,
Water and the Environment



BOOSTING DIAGNOSTIC CAPACITY FOR PLANT PRODUCTION
INDUSTRIES

PROGRAM MAP

(a Rural R&D for Profit program funded project)

Objective 5.1 – Building industry skills
and capability (**industry capability**)

This will be achieved by lifting
capability through:-



Industry Network Training

Tasmanian NPPPs+HPPs - TAS
DPIPWE (Jamie Davies)
ASSCT Presentation plus - SRA
(Kevin Powell)
NT Farmers Federation - NT DPIR
(Maxine Piggott)
(Act 5.1.a)

Workshops & ID Resources

In-field technology -AGVIC
(Fiona Constable)
ISPY – SARDI (Danièle Giblot-
Ducray)
HPPP IDs - CESAR (Jess Lye)
Tasmanian HPPPs - TAS
DPIPWE (Jamie Davies)
(Act 5.1.b)

Evaluate Diagnostic Testing
Programs

QDAF (Paul Campbell)
AGVIC (Fiona Constable)
NT DPIR (Maxine Piggott)
PFR NZ (Simon Bulman)
TAS DPIPWE (Jamie Davies)
SARDI (Danièle Giblot-Ducray)
NSW DPI (Toni Chapman)
(Act 5.1.d)

Assay Development

Aphid/Mealy bug LAMP protocols - WAAA (Monica Kehoe)
Begomovirus field + Ploverovirus rapid protocols - QDAF (Paul Campbell)
Cyst Nematode genome sequenced + Grapevine high throughput +
Fusarium Oxysporum LAMP + Xanthomonas Fragariae high throughput +
LAMP protocols - AGVIC (Fiona Constable)
Fusarium Mangiferae LAMP protocol - NT DPIR (Maxine Piggott)
Xanthomonas minION protocol - PFR NZ (Simon Bulman)
Yellow Canopy Syndrome spectral signature protocol -SRA (Kevin Powell)
Cyst Nematode TaqMan MGB protocol - SARDI (Danièle Giblot-Ducray)
(Act 5.1.c)

Objective 5.2 – Increasing diagnostic options for
rapid detection (**rapid detection**)

This will be achieved by enhanced
tools through:-



Early Detection Tools

Generic virus+rapid vector ID lab protocols +
amplicon sequencing – WAAA (Monica Kehoe)
SWD trap design + endemic cyst nematodes LAMP
test - AGVIC (Fiona Constable)
HPPP eDNA+genetic variant protocol (stop/go) -
CESAR (Jess Lye)
Xanthomonas MinION protocol – PFR NZ (Simon
Bulman)
Xanthomonas citri cg/wgMLST or equivalent – NSW
DPI (Toni Chapman)
Xanthomonas exotic bacteria assay + lab sample
SOPs for NFBSP – QDAF (Paul Campbell)
(Act 5.2.a)

Reference Collections

National Insect (Nematode) Collection – CSIRO
(Mike Hodda)
National Collection of Fungi -AGVIC (Fiona
Constable)
WINC + Emu database – SARDI (Danièle
Giblot-Ducray)
Tasmanian Collection Holdings- TAS DPIPWE
(Jamie Davies)
(Act 5.2.c)

Geographical Origin
Protocols

Cross sectoral HPPPs -
Lincoln NZ (Karen
Armstrong)
(Act 5.2.a)

NDPs Developed/Finalised

Tropilaela Mite – TAS DPIPWE (Jamie Davies)
Cereal+Potato cyst nematodes & Grapevine Red
Blotch & Xanthomonas Fragariae & SWD - AGVIC
(Fiona Constable)
Xanthomonas citri subsp Malvacearum – NSW DPI
(Toni Chapman) & PFR NZ (Simon Bulman)
Cotton Leaf Roll Dwarf & Begomovirus – cotton &
Xanthomonas Axonopodis pu allii & Cotton Leaf
Curl – QDAF (Paul Campbell)
Cryptotermes (drywood termite) – NT DPIR (Maxine
Piggott)
Bursaphelenchus (pine-wood & red ring
nematodes) & cyst nematodes – CSIRO (Mike
Hodda)
Exotic cyst nematodes -SARDI (Danièle Giblot-
Ducray)
(Act 5.2.d)

SPHD Adoption/Endorsement of NDPs

Tropilaela Mite – TAS DPIPWE (Jamie Davies)
Cereal+Potato cyst nematodes & Grapevine Red
Blotch & Xanthomonas Fragariae & SWD - AGVIC
(Fiona Constable)
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Piggott)
Bursaphelenchus (pine-wood & red ring
nematodes) & exotic cyst nematodes – CSIRO (Mike
Hodda)
Exotic cyst nematodes -SARDI (Danièle Giblot-
Ducray)
Aphids/Mealy Bug – WAAA (Monica Kehoe)
(Act 5.2.f)

Objective 5.2 – Increasing
diagnostic options for rapid
detection (**rapid detection**)

This will be achieved by
enhanced tools through:-



Train against NDPs

HPPP workshops – SARDI
(Danièle Giblot-Ducray)
Tasmanian HPPP presentations
& factsheets - TAS DPIPWE
(Jamie Davies)
(Act 5.3.a)

Incursion Scenario

TBC 3-5 day workshop – PHA
(Natalie O'Donnell)
(Act 5.3.b)

Blitzes

Surveillance + response – NT
DPIR (Maxine Piggott)
Surveys – QDAF (Paul
Campbell)
Bioblitzes & sample collection
kits – TAS DPIPWE (Jamie
Davies)
AUSPestCheck data entered –
SARDI (Danièle Giblot-Ducray)
(Act 5.3.c)

Proficiency Testing

Ring test assays - QDAF (Paul
Campbell) + NSW DPI (Toni
Chapman) + AGVIC (Fiona
Constable) + PFR NZ (Simon
Bulman)
3-4 national labs – TAS
DPIPWE (Jamie Davies)
Sequencing dry lab – NSW DPI
(Toni Chapman)
(Act 5.3.c)

Case study: Using *Xanthomonas citri* subsp. *Malvacearum* as a model organism for increasing bacterial diagnostic capacity



Department of
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NSW DPI:

- Using *Xanthomonas* as a model organism for the development of new techniques for identification
- Determine current diversity of *Xanthomonas* in Australia
- Human capabilities through employment of PhD student

Plant and Food Research NZ:

- Develop diagnostic protocols for *Xanthomonas*

Cross collaboration is key!



Bacterial blight of cotton

Source: Clemson University – USDA Cooperative Extension Slide Series, Bugwood.org



Case study: Nematode diagnostics



CSIRO:

- Developing human capabilities (succession planning)
- Developing collections of local and exotic nematodes
- Updating national diagnostic protocols and producing information resources



SARDI:

- High throughput diagnostics for exotic cyst nematodes including development of molecular assays for two species
- Tests are efficient and effective



Jobs,
Precincts
and Regions

Agriculture Victoria:

- Build capability in cyst nematode taxonomy (*Heterodera*) – PhD student
- Utilise suitable genomics approaches to identify suitable molecular markers for species identification
- Develop diagnostic tests to detect key cyst nematodes in soil



How does this fit within our community?

- BEC connects extension professionals.
- Significant investment into diagnostics through this project.
- Sharing diagnostics with industry is critical.
- Written communications, field days, pest and disease management workshops, webinars – endless opportunities.

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Collaborating to ensure exotic pest diagnostic capacity

14 SEP 2021

Giving Australian agr diagnostics a boost



While dealing with bacterial blight is a thing of the past for Australian cotton growers due to resistant cultivars, the threat of an incursion from exotic, more harmful strains remains.

CRDC is supporting research to prepare the Australian industry for a possible incursion, as part of a cross industry partnership. A key focus is on the *Xanthomonas citri* subsp. *Melanoconum*, the bacteria which causes bacterial blight in cotton and diseases such as citrus canker in fruit trees. There are several described races and distribution varies between countries. Bacterial blight of cotton was first reported in the US in Alabama in 1891 and is now widely distributed. Although strains of bacterial blight are already present



Symptoms of bacterial blight start as small water-soaked lesions on the leaves. As the disease progresses the lesions turn black and pupery with premature defoliation of the leaf occurring. If these symptoms are detected contact the Emergency Plant Pest hotline on 1800 084 881.

diagnosed and identified, how we develop diagnostic assays, and the development of National Diagnostic Protocols through the inclusion of new technologies for faster turnaround times.

The researchers will use genome sequences to develop diagnostic assays for the target pathogen *X. citri* subsp. *Melanoconum*, using these new diagnostics to update the National Diagnostic Protocol for bacterial blight of cotton, and update the identities of the Australian reference collection of *Xanthomonas*.

"The benefit of this project is that *Xanthomonas* will be used as the model organism as we have readily available genome sequences," Toni said.

"All the methodologies developed and knowledge gained can be translated to other phytopathogens including, *Pseudomonas*, *Xylella*, *Agrobacterium* and *Dickeya*."

CRDC R&D Manager Susan Maas says if a blight-like pathogen is detected in



Acknowledgements

Funding body



Project lead



RDCs



Wine Australia

Service Providers



Department of Primary Industries and Regional Development



Department of Primary Industries



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